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# Osteoarthritis and Cartilage



## Review

## Mechanics and biology in intervertebral disc degeneration: a vicious circle



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### SUMMARY

Intervertebral disc degeneration is a major cause of low back pain. Despite its long history and large socio-economical impact in western societies, the initiation and progress of disc degeneration is not well understood and a generic disease model is lacking. In literature, mechanics and biology have both been implicated as the predominant inductive cause; here we argue that they are interconnected and amplify each other. This view is supported by the growing awareness that cellular physiology is strongly affected by mechanical loading. We propose a vicious circle of mechanical overloading, catabolic cell response, and degeneration of the water-binding extracellular matrix. Rather than simplifying the disease, the model illustrates the complexity of disc degeneration, because all factors are interrelated. It may however solve some of the controversy in the field, because the vicious circle can be entered at any point, eventually leading to the same pathology. The proposed disease model explains the comparable efficacy of very different animal models of disc degeneration, but also helps to consider the consequences of therapeutic interventions, either at the cellular, material or mechanical level.

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## Introduction

Low back pain is a top-3 cause of disability in developed countries, and the number of people affected is increasing worldwide<sup>1</sup>. Up to 40% of adult persons in the United States report low back pain in the preceding 3 months, and with 20–33% of patients being unable to work, the disease has a major socio-economic impact<sup>2,3</sup>. In the Netherlands, recent policy changes in the management of low back pain have decreased expenditure, but the total costs are still 216 euro's per capita annually<sup>3</sup>. Prevention and therapeutic intervention is hampered because the veritable cause

of low back pain remains unclear; however, a correlation with intervertebral disc degeneration has been documented<sup>2,4–10</sup>. Unfortunately, the aetiology of intervertebral disc degeneration is as obscure as the cause of low back pain, and the current consensus is that it is “multi-factorial”. Numerous changes in disc morphology and physiology have been described, but these alterations have not yet lead to a widely accepted disease model. The lack of an accepted explanatory model limits the understanding of this disabling disease, and hampers the development of effective therapies.

One of the issues to be resolved is the order and causal relationship of the biological and biomechanical alterations that occur in intervertebral disc degeneration. Some authors hypothesize that disc degeneration originates from biomechanical wear and tear<sup>11–13</sup>. Other authors focus on the disturbance of physiological cellular behaviour, mainly based on a loss of nutrition<sup>14–17</sup>, but recently pathogens have been implicated as well<sup>18</sup>. However, these two viewpoints do not exclude each other, and it is conceivable that different pathological processes cause the same disease, equivalent to the etiological disease model of diabetes mellitus with subtypes 1 and 2. In fact, the dichotomy between biology and mechanics

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currently seen in the field may be unnecessary, as it has long been recognized that cellular physiology is affected by its mechanical environment<sup>19,20</sup>. This relationship, known as mechanobiology<sup>21</sup>, has recently also been established for the intervertebral disc<sup>22–24</sup>, and is deemed instrumental in developing intervertebral disc degeneration<sup>25–29</sup>.

Similar to developing wrinkles in the skin, degeneration of the intervertebral disc is part of normal aging. In analogy to this, the painful degenerative disc disease<sup>30</sup> has been likened to accelerated aging of the disc<sup>31</sup>. As such, it is important to realize similar processes occur in aging and degeneration alike, and a clear discrimination between the two is difficult. Additionally, there is a strong genetic basis for intervertebral disc degeneration<sup>32,33</sup>, because genetic information determines cellular behaviour and structural integrity of produced extracellular matrix. Therefore, polymorphisms in genes such as COL1A1 and or ADAMTS5 are a risk factor for developing degeneration at a younger age<sup>34,35</sup>. Nevertheless, neither age nor genetic make-up can be remedied; therefore, it is more relevant to look at what underlying processes are involved in order to halt or reverse intervertebral disc degeneration.

In this paper, we present a contemporary disease model of intervertebral disc degeneration. While this model can not explain low back pain, the development of a disease model is essential in identifying lapses in knowledge and development of therapies for associated intervertebral disc degeneration. Our disease model is based on the changes that occur in the nucleus pulposus, and is in the form of a positive feedback loop involving cells, extracellular matrix, and biomechanics (Fig. 1). Novel in this model are the mechanobiological cues that close the loop from biomechanics to cells, and involve a shift from hydrostatic stress to shear stress in the nucleus pulposus. In order to demonstrate that most common risk factors for developing intervertebral disc degeneration can initiate the positive feedback loop, we additionally apply the interactions in this model to human epidemiology, and observations in the different animal models for disc degeneration. The deliberation of the model will be preceded by a short introduction to the functional anatomy of the intervertebral disc and the changes of structures with degeneration.

### The intervertebral disc and its anatomical structures in health and degeneration

Intervertebral discs are embedded between the vertebrae and provide flexibility to the spine. They consist of three anatomical parts: the nucleus pulposus, the annulus fibrosus, and the cartilaginous endplates. The nucleus is the core of the intervertebral disc, and is surrounded by the lamellae of the annulus fibrosus. Cranially and caudally the endplates limit the intervertebral disc, and form the anchoring into the vertebral bodies. Disc degeneration is associated with changes in all these anatomical structures.

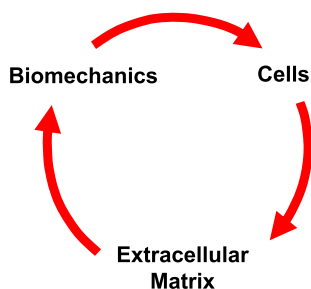


Fig. 1. Concept of the degenerative circle of intervertebral disc degeneration.

These alterations have been extensively reviewed in numerous papers<sup>15,30,35–42</sup>, hence only short summary of the nucleus', annulus' and endplates' structure in a normal and a degenerated intervertebral disc will be provided.

A healthy nucleus pulposus is a gel-like, highly hydrated tissue, rich in proteoglycans<sup>43</sup>. The healthy nucleus pulposus generates an intradiscal pressure which separates the two vertebrae, tensions the annulus fibrosus, and distributes pressure evenly over the two adjacent endplates<sup>41,44–46</sup>. A degenerated nucleus pulposus is an unorganized fibrous tissue which has largely lost its capacity to bind water under compression<sup>43</sup>. Therefore, the pressure in the nucleus pulposus is dwindling<sup>47</sup>, and disc height is lost<sup>45,46</sup>. Overall, the nucleus undergoes the highest degree of remodelling during intervertebral disc degeneration<sup>44,48</sup>.

A healthy annulus fibrosus is a highly organized fibrous structure. It consists of ~20 concentric lamellae of alternating oblique collagen fibres interspersed with proteoglycans<sup>49,50</sup>. The collagen fibres are tensioned by intradiscal pressure through two mechanisms: direct radial pressure from the nucleus pulposus, and cranial-caudal stretch from the separation of the two endplates<sup>41,44,45</sup>. Due to a loss of intradiscal pressure, the annulus fibrosus of a degenerated intervertebral disc deforms by in- and out-ward bulging and buckling<sup>45,51,52</sup>, and shows progressive increase of structural defects such as: rim lesions, de-lamination and radial fissures<sup>30,41,49</sup>. Remarkably, despite these structural changes, there is hardly any loss of tensile strength<sup>53,54</sup>; however, hydraulic permeability changes from anisotropy favouring the radial direction to isotropy<sup>55,56</sup>, which could affect the build-up of intradiscal pressure.

Healthy vertebral endplates are of uniform thickness, do not bulge into the vertebrae and appear as homogeneous hyaline cartilage<sup>51,57</sup>. With intervertebral disc degeneration, there is an increase in microscopic and macroscopic damage to the endplate<sup>37,58</sup>. Additionally, there is a marked increase in sclerosis of the subchondral bone<sup>59–61</sup>, similar to degenerated cartilage<sup>62</sup>. Changes in endplate and subchondral bone morphology (e.g., fractures or endplate sclerosis) have also been implicated as preceding intervertebral disc generation (by decompression of the nucleus<sup>12,31,63,64</sup> or impairment of nutrition<sup>16,60,65</sup>, respectively). Overall, the endplate can be deemed an important part of the intervertebral disc, because damage to the endplate is strongly related to both intervertebral disc degeneration and low back pain<sup>5,58,66,67</sup>.

Overall, a degenerated intervertebral disc differs from a non-degenerated intervertebral disc in that there is a marked loss of disc height, a fibrous dehydrated nucleus, in-ward and out-ward buckling of annulus fibres, extensive endplate damage, and sclerosis of the subchondral bone.

### Degeneration of the intervertebral disc; an interaction between cells, extracellular matrix, and biomechanics

The nucleus pulposus radiographically shows the most extensive changes in intervertebral disc degeneration<sup>44,48,68</sup>, and it is therefore the most thoroughly investigated. Both the annulus fibrosus and cartilaginous endplates have received attention in their relationship with intervertebral disc degeneration; however, changes in these structures are less well documented<sup>37,51</sup>. Therefore, this section will focus on the changes in the nucleus pulposus, followed by a short summary of the effect of nucleus degeneration on the annulus and endplates, and vice versa. We will discuss the cells in the nucleus pulposus and their interaction with the surrounding matrix; the effect of the shift of matrix composition on the biomechanical behaviour; and the subsequent effect of biomechanical stresses on cellular physiology. This will show the progressive nature of intervertebral disc degeneration to be a

positive feedback loop as shown in its basic conceptual form (Fig. 1).

#### *Cells: from notochordal cells to nuclear chondrocytes*

In the human nucleus pulposus, notochordal cells that are present from the early embryonic formation of the intervertebral disc<sup>69,70</sup> show a gradual transition towards chondrocyte-like cells in the first decade of life<sup>37,38</sup>. Recently murine fate mapping studies demonstrated that the mature chondrocyte-like cells in the nucleus pulposus cells are derived from the embryonic notochord<sup>71,72</sup>. These mature nuclear chondrocytes produce collagen type I, but reduced amounts of water-attracting proteoglycans and collagen type II<sup>42</sup>. Thus, the transition of the cell population in the nucleus pulposus from predominantly notochordal cells to chondrocyte-like cells leads to a decrease in proteoglycan synthesis and therefore affects the potential of the nucleus pulposus to maintain its structure and composition<sup>73,74</sup>.

#### *Cells – extracellular matrix: from anabolism to catabolism*

In the degenerating intervertebral disc, there is a progressive increase in the expression of inflammatory cytokines like IL-1 and TNF $\alpha$ <sup>75–77</sup>. These cytokines, expressed by nucleus cells, up-regulate matrix remodelling involved in intervertebral disc degeneration<sup>75,78</sup>. Matrix remodelling by the nucleus cells is mainly mediated by two families of enzymes: Matrix Metallo Proteinases (MMP) and A Disintegrin And Metalloproteinases with Thrombospondin Motifs proteins (ADAM-TS)<sup>36,79</sup>. Some non-proteolytic degradation is also present due to glycation<sup>80</sup>. In later stages of disc degeneration, inflammatory cytokines also enhance neurovascular in-growth and pain response<sup>75,78</sup>. Altogether, there is a progressive reduction in the expression of proteoglycans and collagen type II genes with increasing degeneration<sup>36,42,48,81</sup>. Simultaneously, collagen type I expression is increased, which indicates a change in matrix stresses<sup>82</sup>.

#### *Extracellular matrix: from proteoglycans to collagen type I*

The nucleus pulposus extracellular matrix consists of proteoglycans and collagens, and aggrecan is by far the most abundant proteoglycan in the nucleus<sup>43</sup>. Proteoglycans have a negative charge, which causes an osmotic pressure of 420–450 mOsm<sup>83</sup>. This osmotic pressure attracts and binds water to the extracellular matrix. In degeneration, aggrecan is cleaved from the hyaluronic acid backbone<sup>84,85</sup>. Cleaved aggrecan fractions do not aggregate<sup>43</sup>, making them less effective in binding water. Furthermore, there is a shift of predominantly collagen type II to collagen type I in the nucleus<sup>42</sup>. Overall, the biochemical content of the extracellular matrix changes from predominantly proteoglycans and collagen type II to a more fibrous tissue consisting primarily of collagen type I, resulting in a loss of water-binding potential.

#### *Extracellular matrix – biomechanics: a reduction in intradiscal pressure*

In healthy discs, the negative charge of the proteoglycans generates an osmotic potential, which is translated into a biomechanical hydrostatic pressure through the attraction of water. This intradiscal pressure is approximately 0.1–0.24 MPa when lying supine, and increases linearly with loading of the disc<sup>47,86–88</sup>, up to more than 2.0 MPa<sup>86</sup>. The quantity of bound water can vary, which changes the intrinsic intradiscal pressure<sup>89</sup>. In healthy discs, this decrease or increase of bound water is due to poro-elastic fluid flow upon loading or unloading of the disc, respectively<sup>46,90</sup>. In

degenerating discs, the increased fragmentation of aggrecan reduces its effective negative charge, which decreases intradiscal pressure<sup>47</sup> and the ability to retain water under compressive forces<sup>91</sup>, which is reflected in the reduction of disc height<sup>44,46</sup>. The effect of a reduction of collagen type II and an increase of collagen type I on the biomechanical function of the nucleus matrix is unknown. However, as collagen type II is more compliant than collagen type I, an increase of nuclear shear stresses is expected.

#### *Biomechanics: from hydrostatic pressure to shear stress*

Intradiscal pressure is essential for the maintenance of biomechanical behaviour of the intervertebral disc. Intradiscal pressure tensions annulus fibres, and supports the endplate, and as such is the main determinant of disc height and stiffness in axial compression<sup>45,46</sup>. In degenerated intervertebral discs, disc height and axial compliance are reduced, and radial bulge is increased<sup>45,46,92</sup>. Another effect of the reduced intradiscal pressure in the intervertebral disc is the disturbed stress distribution found in degenerated discs<sup>30,93</sup>. This disturbance in stress distribution generates stress concentrations, which increases the risk of endplate fractures or Schmorl's nodes, which are increasingly seen with disc degeneration<sup>66</sup>.

A reduction in intradiscal pressure leads to increased shear stresses in both the nucleus pulposus and the annulus fibrosus upon axial compression of the spine<sup>17,90</sup>. Due to loss of tension in the annulus fibrosus, motion segments with reduced intradiscal pressure also have an enlarged neutral zone in shear, bending, and torsion<sup>41,45,94–98</sup>. The resultant changes in bending and torsion behaviour of the motion segment may further increase shear stresses in the nucleus and remodelling of the extracellular matrix. Thus, the reduction of intradiscal pressure reduces disc height; increases stress concentrations within the disc; and increases shear forces in the nucleus.

#### *Biomechanics – cells: a change in matrix stresses alters cellular physiology*

The concept that the mechanical environment of cells is important for cell function is not new. In 1862, Hueter and Volkmann independently hypothesized that mechanical stimuli directly influence cellular function and matrix synthesis in bone and joints due to local differences in tension and pressure<sup>19,20</sup>. Today, the effect of biomechanical forces on cellular function is known as mechanobiology<sup>11,21,25,28</sup>. Several research groups have shown that a distinct compressive force on the spinal motion segment, both *in vivo* and *ex vivo*, can cause catabolic, anabolic and inflammatory cell responses in the intervertebral disc<sup>23,26,99–102</sup>. Also the temporal characteristics of loading are important as cyclic loading has been shown to be beneficial as opposed to static loading<sup>22,99,103,104</sup>. As a result, the relationship between mechanical behaviour and cell function is argued to be a pivotal component of intervertebral disc function and dysfunction<sup>25,27,28,105</sup>.

Cells throughout the intervertebral disc respond to changes in hydrostatic pressure. In the nucleus, the proteoglycan production at 0.3 MPa is roughly 20% higher than at 0.1 MPa<sup>106,107</sup>. Additionally, MMP-3 production is reduced, and tissue inhibitor of metalloproteinases-1 (TIMP) production is increased<sup>106,107</sup>, which reduces remodelling of the extracellular matrix. This pressure sensing mechanism of nucleus cells appears to be impaired in cells from degenerated discs as they respond less anabolic to physiologic intradiscal pressure<sup>108</sup>. Cells also respond to the osmotic pressure of the extracellular matrix, with an optimum proteoglycan production at pressures between 400 and 500 mOsm, and a reduced synthesis of aggrecan with declining or increasing osmotic

pressure<sup>83,109–111</sup>. A decline in osmotic pressure increases MMP-3 production<sup>111</sup>, and precludes hypertrophy of the normally hyperosmotic nuclear chondrocytes<sup>17,112</sup>. Thus, in degenerating intervertebral discs, the drop in intradiscal and osmotic pressure will reduce the anabolic stimulus and increase catabolic stimuli to the nuclear chondrocytes.

The shift of hydrostatic pressure to shear stresses in the intervertebral disc has a distinct mechanobiological effect on the nuclear chondrocytes<sup>90,113,114</sup>. Similar to other load-bearing tissues like cartilage and bone, the increase in shear stress will initiate the formation of a fibrous tissue, rich in collagen type I<sup>25,82,114,115</sup>. Furthermore, increased shear stress increases the production of nitric oxide by chondrocytes<sup>114</sup>. Nitric oxide is a reactive oxygen metabolite that reduces proteoglycan production, and increases apoptosis in cartilage and in the intervertebral disc<sup>114,116,117</sup>. Thus, reduction of intradiscal pressure increases shear stresses in the nucleus, and both may accelerate degeneration in the intervertebral disc.

#### *Nucleus homeostasis depends on endplate and annulus integrity*

Although we have focussed on the nucleus pulposus in this section, the homeostasis of the nucleus is dependent on the confines of a functional annulus and intact endplates<sup>109,118</sup>. If damage to either of these structures occurs, the nucleus is decompressed<sup>12,119</sup>, and exposed to inflammatory cells from outside the disc<sup>78</sup>. Both these effects will result in the degenerative cascade described above. Conversely, if the nucleus is degenerated, this will also affect the annulus and the endplates. In the annulus, the reduction of intradiscal pressure will reduce tension in annulus fibres<sup>45</sup> and increase in- and out-ward bulging<sup>12,52</sup>. This bulging can increase shear forces between laminae<sup>120</sup>, leading to delamination of the translamellar bridges<sup>121</sup>, and consecutive risk of tears<sup>52</sup>. In the endplates, the loss of annulus tension and the reduced stress distribution by the nucleus will alter the biomechanical stresses on the endplates<sup>12,44</sup>, which may be the cause of endplate sclerosis, fractures, or Schmorl's nodes<sup>66</sup>.

In summary, the interaction of cells, extracellular matrix and biomechanical stress is instrumental in homeostasis of the

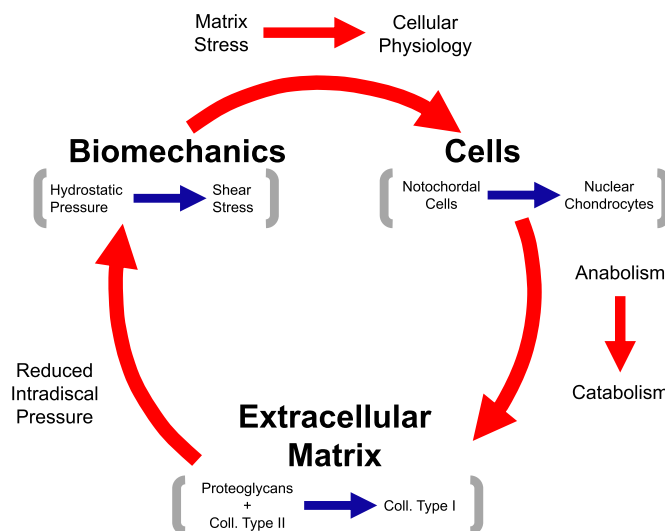
intervertebral disc. In intervertebral disc degeneration this balance is disturbed. If the cells do not receive the proper mechanical and chemical cues they will stop producing, or even start degrading proteoglycans. A reduction in proteoglycans will lead to a drop of the intradiscal pressure, which will alter the biomechanical stresses on the cells. From this, one can deduce a positive feedback loop of intervertebral disc degeneration, which contains cells, extracellular matrix, and biomechanics: the degenerative circle (Fig. 2).

#### **Application of the degenerative circle**

The degenerative circle illustrates the progressive nature of intervertebral disc degeneration, but can also explain the different aetiologies of intervertebral disc degeneration. In this section, we investigate the application of the degenerative circle in understanding human epidemiology and animal models for intervertebral disc degeneration. In human epidemiology, aberrant biomechanics (e.g., frequent lifting<sup>122</sup>); chemical stress to cells (e.g., smoking<sup>123</sup>); or damage to the extracellular matrix (e.g., discography<sup>124</sup>); all lead to intervertebral disc degeneration. Additionally, induction of intervertebral disc degeneration in animal models can be effectuated through: altered disc biomechanics, changes to cell physiology, and damage to the nucleus, annulus, or endplates. This section will provide examples of the initiation of degeneration through each of the three domains, i.e., biomechanics, cells, and extracellular matrix. By applying the model from different angles, we aim to infer the generic nature of the degenerative circle, as all discussed examples of human epidemiological occurrence of disc degeneration and animal models apparently lead to a similar degeneration of the intervertebral disc (Fig. 3). To illustrate the independence of starting at a specific point in the circle, we start this section by discussing biomechanics.

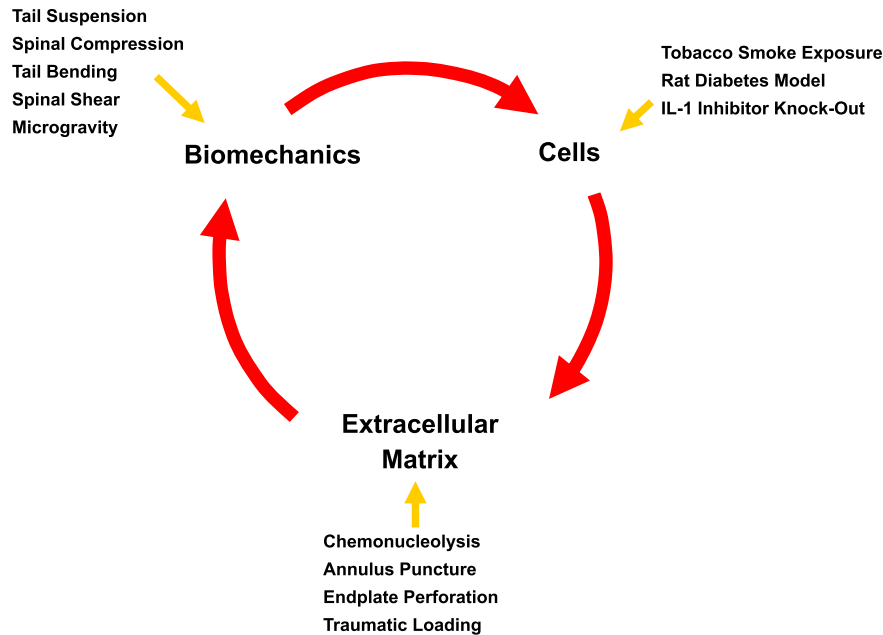
#### *Biomechanics: induction of degeneration*

In literature the biomechanical “wear and tear” has long been thought to be a major cause of intervertebral disc degeneration<sup>32,125</sup>, mainly because low back pain and degeneration occur, more frequently than in the general population, in manual labour



**Fig. 2.** The degenerative circle of intervertebral disc degeneration. Homeostasis of the intervertebral disc is dependent on the interaction of cells, extracellular matrix and biomechanical stress. If this balance is disturbed, the cells stop producing proteoglycans, this will give a reduction in hydrostatic pressure and increase shear forces on the cells. An increase of shear forces further decreases the production of proteoglycans, leading to progressive degeneration.





**Fig. 3.** The efficacy of animal models applied to the degenerative circle. Intervertebral disc degeneration can be induced through any of the three main elements of the degenerative circle, which further indicates a positive feedback loop.

workers<sup>7</sup>, machine drivers<sup>7</sup>, soldiers carrying loads<sup>126</sup>, but also in elite athletes<sup>127–129</sup>. Interestingly, all astronauts experience low back pain upon the exposure to microgravity, and on their re-entry<sup>130,131</sup>, which both may be caused by over-pressurization of the nucleus. Although genetic research has nuanced the role of biomechanical factors in intervertebral disc degeneration<sup>32</sup>, there still is a link between high loading on the low back and both intervertebral disc degeneration and low back pain<sup>5,7,58,132–134</sup>.

An abundance of animal models uses altered biomechanics to induce intervertebral disc degeneration, including: Tail suspension/Hind leg unloading<sup>135,136</sup>; Tail or spinal compression<sup>26,137–140</sup>; tail bending<sup>141–143</sup>; spinal shear stress<sup>113</sup>, and microgravity<sup>144,145</sup>. These models show that although the intervertebral disc is left intact, the altered biomechanical load leads to a catabolic cell reaction and remodelling of the intervertebral disc matrix over time. Apparently, in animal models, it does not matter whether the disc is overloaded, unloaded or aberrantly loaded: altering the biomechanical environment of the intervertebral disc induces a catabolic cell reaction with detrimental effects on the extracellular matrix.

#### Cells: induction of degeneration

One of the most influential paradigms on intervertebral disc degeneration is that a reduction in nutrition of disc cells leads to a catabolic shift<sup>15,16,35,65</sup>. The hypothesis is that this is due to the sclerosis of the endplates, which limits endplate pores and subsequent vascular supply<sup>60</sup>. It has been established that diffusion into the disc changes with progression of intervertebral disc degeneration<sup>146</sup>. However, the origin of endplate sclerosis should be further elucidated to determine whether endplate sclerosis is in fact the cause, or merely an effect of degeneration due to altered biomechanical stresses in the endplates. Other risk factors like smoking<sup>32,123</sup> and diabetes mellitus<sup>147</sup>, most likely induce disc degeneration by their effect on cellular physiology. Interestingly, these risk factors may also affect the nutrition of the nuclear chondrocytes by their detrimental effects on microcirculation<sup>35,148–151</sup>. Additional to the effects of nutrition, low-grade

infection could possibly trigger the cells to degrade the matrix of the intervertebral disc<sup>18,152</sup>, similar to arthritic diseases<sup>153–155</sup>.

Intervertebral disc degeneration is found in mice which are exposed to tobacco smoke<sup>156,157</sup>, and in rat models for diabetes<sup>158,159</sup>. The exact pathophysiological pathway is not clear, but some information may be gleaned from these experimental models. Disc degeneration in tobacco smoke models is not mediated by genotoxic DNA damage, but by an alteration of cell physiology<sup>160</sup>. This may be caused by the increase of the nitric oxide concentration in the blood, which reduces proteoglycan synthesis<sup>116</sup>. In diabetes models, hyperglycaemia could play a role, either by a direct effect on nucleus cells<sup>161,162</sup>, glycation reactions with aggrecan<sup>80</sup>, or by the increase of the osmotic value of the blood. However, in both models a biomechanical effect cannot be excluded. In smoke models, the vertebral bodies show a marked increase in porosity, which reduces the structural integrity. In diabetes models, the overweight may induce overloading. Again, it could also be the negative effect on the microcirculation that both smoking and diabetes mellitus have in humans; however, to our knowledge, the effect of smoking or diabetes on endplate microcirculation has not yet been investigated in animal models.

Evidence for induction of intervertebral disc degeneration through a catabolic shift in cells is not well established in animal models, but there is evidence from IL-1-inhibitor knock-out mice (IL-1 $\alpha$ <sup>-/-</sup>) that raised levels of IL-1 $\beta$  coincide with intervertebral disc degeneration after 55 days<sup>163</sup>. *Ex vivo*, injection of MMP-3, ADAM-TS4 or HTRA-1 showed little effect on catabolic gene expression after 8 days<sup>164</sup>; however, TNF $\alpha$  addition to the culture medium has been shown to have a persistent catabolic effect on disc cells up to 21 days<sup>76</sup>. Infectious processes that induce intervertebral disc degeneration have to our knowledge not been investigated in animal models.

#### Extracellular matrix: induction of degeneration

Herniation of the nucleus<sup>165</sup>, puncture of the annulus<sup>124</sup>, or endplate fracture<sup>63,166</sup> are associated with the long-term risk of disc degeneration in humans. This damage can be induced through a

single traumatic overload<sup>166,167</sup>, which can damage the extracellular matrix, both macroscopically<sup>12,165</sup> and microscopically<sup>168</sup>. This results in a loss of intradiscal pressure<sup>119,166,169</sup>, and significantly elevated levels of interleukin (IL)-5, IL-6, IL-7, IL-8, MCP-2, GRO $\alpha$ , MIG and NGF<sup>166,167</sup>. Interestingly, it appears that the damage to the matrix, either endplate or annulus, is essential for developing intervertebral disc degeneration, rather than simply the absorption of a distinct amount of energy<sup>170</sup>. The induction of degeneration then occurs by decompression of the nucleus<sup>119,166,169</sup>, exposure of nucleus cells to matrix fragments<sup>171</sup>, response to neurotrophic and angiotrophic factors<sup>78</sup>, or a combination thereof. This illustrates that within the domain of extracellular matrix there are different pathways into the degenerative circle<sup>31</sup>, which appear to depend upon decompression and exposure of the nucleus<sup>12,166</sup>.

In animal models, damage to the extracellular matrix is the most commonly used method of induction of intervertebral disc degeneration. Whether the damage is done by chemo-nucleolysis<sup>172–175</sup>, annulus puncture<sup>176–179</sup> or endplate perforation<sup>169,180</sup>, progressive degenerative disc degeneration is seen. Unfortunately, a comparison of the chronological order of cellular and biomechanical changes between these different methods of degeneration induction has not been performed. However, it has been established that pressure drop<sup>181</sup> and the expression of catabolic agents<sup>170,182,183</sup> occur both *ex vivo* and *in vivo*. Interestingly, it has been shown that disc stress distributions in the IVD are influenced more by damage to the endplate than by injuries to the outer annulus<sup>64</sup>, but again, direct comparison of the differences in cellular changes between these two pathways has not yet been performed.

In summary, both in human epidemiology and in animal models there is evidence for numerous pathways towards progressive disc degeneration. This is important because it illustrates why intervertebral disc degeneration has been called multi-factorial. The degenerative circle can explain most of the common risk factors for intervertebral disc disease, and the progressive nature of degenerative disc disease.

## Discussion

In this paper we propose a model for intervertebral disc degeneration: the degenerative circle. This model is based on the most prominent alterations that occur in the nucleus pulposus in intervertebral disc disease, and consists of a positive feedback loop involving cells, extracellular matrix, and biomechanics. Additionally, this paper aims to provide insights into the pathways into the degenerative circle based on human epidemiology and animal models for intervertebral disc degeneration.

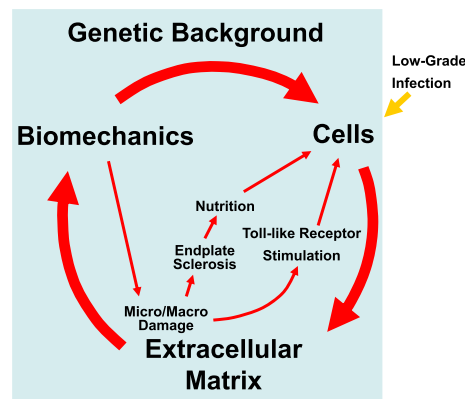
Both Adams *et al.*<sup>11</sup> and Colombini *et al.*<sup>184</sup> have proposed pathophysiological models for intervertebral disc degeneration that include some of the relations of the degenerative circle. The model of Adams *et al.* focuses on structural damage to the extracellular matrix and is progressive due to a frustrated cellular healing response, mainly because of a drop of intradiscal pressure. Their model thus differs in two fundamental ways: firstly, it only allows for disc degeneration to occur upon damage to the extracellular matrix. Secondly, in their model, mechanobiological cues are limited to a decrease in intradiscal pressure, and do not include an increase in shear stresses. However, this seems crucial for the breakdown of aggrecan, and the transdifferentiation to collagen type 1 producing cells. The model of Colombini *et al.* regards chronic abnormal load as the main cause of intervertebral disc degeneration; they state that this will lead to a catabolic cell response, and consecutively an altered matrix. There are similarities with the degenerative circle; however, again their model differs in crucial ways: their model does not allow for aberrant cell

physiology or damage to the extracellular matrix to induce disc degeneration, nor does it elaborate on how the catabolic cell response is induced. Furthermore, their model does not stress the progressive nature through a positive feedback loop. The degenerative circle thus presents a more complete view of intervertebral disc degeneration as it allows for multiple ways of induction of intervertebral disc degeneration, illustrates the progressive nature through a positive feedback loop, and is the first to elaborate on the mechanobiological cues that play a role in intervertebral disc degeneration.

The degenerative circle is a simple model. It provides a practical tool for clarifying the complex interactions of intervertebral disc disease to patients, medical students, and clinicians. Additionally, this model stresses the importance of the interaction between cells, extracellular matrix and biomechanical behaviour, and illustrates that all are important in intervertebral disc degeneration. This is essential because all three domains and their interactions need to be considered if we want to reverse or halt the degenerative process. However, the simple elegance of the degenerative circle has intrinsic shortcomings as it contains some oversimplifications.

Weaknesses in the proposed model include a lack of other feedback mechanisms in intervertebral disc physiology. Clearly, besides a pathway through biomechanical changes, there is also a direct feedback loop from the extracellular matrix to the cells. This is mainly dominated by the osmotic charge of the proteoglycans. Numerous other mechanisms (e.g., endplate sclerosis<sup>65</sup>, the effect of loading on nutrition<sup>185</sup>, low-grade infection<sup>18</sup>, toll-like receptor stimulation<sup>171,186</sup>) could later be added to the model, when their effects are further quantified. It is interesting to note that feedback mechanisms in intervertebral disc degeneration seem to progress the disease rather than halt it, which is remarkable since feedback loops usually poise homeostasis in human physiology. Therefore, more research could be performed to identify anabolic feedback mechanisms in the intervertebral disc. Due to these and other possible effects, the degenerative circle should not be regarded as a definitive model for intervertebral disc degeneration, but rather as the backbone of a more detailed model.

The choice of excluding genetic influences in the degenerative circle was made to simplify the current disease model; however, from the literature it is known that genetic influences play a substantial role in developing intervertebral disc degeneration<sup>32,33</sup>. A loss of matrix integrity due to genetic defects (e.g., Col I, Col IX, VitD, Aggrecan, MMP-3 and MMP-9), has been shown to play a role in the



**Fig. 4.** Additions to the degenerative circle. The degenerative circle is not a final account of intervertebral disc degeneration, possible future additions to the model are included. It is also important to notice that the degenerative circle is drawn upon an individual's genetic background, which probably influences all pathways included in the model.

development of intervertebral disc degeneration<sup>35,187</sup>. Moreover, Rajasekaran *et al.* recently showed that the development of either endplate damage, disc height loss or annulus tears was associated with deficits in specific genes which code for the extracellular matrix of respective intervertebral disc parts<sup>34</sup>. Additionally, there is a clear role for genes in the biomechanical forces (e.g., length, weight<sup>188</sup>), and probably also for the mechanobiological response to biomechanical forces<sup>108</sup>. Therefore, the genetic make-up of single patients can be viewed as the background upon which the degenerative circle is drawn, this is shown in Fig. 4, along with the possible additions to the model.

Animal models studied in a longitudinal manner can be useful in unravelling different cascades of disc degeneration, and understanding the timing of changes. Further understanding of the timing of degenerative changes is essential in the development of prevention and therapies for intervertebral disc degeneration. Chondrodystrophic and non-chondrodystrophic dogs provide an interesting study population as the former develops early disc degeneration whereas the latter only develops disc degeneration at advanced age<sup>189</sup>. This could help distinguishing between aging and degeneration, but also indicate what changes precede, and what changes follow. Furthermore, comparison of the timing of changes between different ways of inducing disc degeneration (e.g., chemonucleolysis vs smoking induced degeneration, or diabetes models vs overloading) may shed a light on whether genetics, environment, or matrix damage indeed provide a similar disc degeneration, or whether if there are differences (and the current disease model should be updated).

The degenerative circle is a model for the multifaceted disc degeneration, but does not explain why some people get low back pain and others do not. It is important to consider that low back pain is a very heterogeneous symptom, in which a discogenic origin is just one of the causes<sup>190</sup>. Discogenic back pain in itself is probably also heterogeneous, depending on damage to innervated parts of the intervertebral disc (i.e., endplate or annulus)<sup>31</sup>. Additionally, the nucleus could also give rise to discogenic back pain, especially upon in-growth of nerve fibres<sup>75,191</sup>. These could be triggered by the increase in inflammatory cytokines<sup>75–77</sup>, which are increasingly produced with degeneration of the hyaluronic acid backbone<sup>171</sup>. This in-growth of nerve fibres should especially be considered in end-stage disc degeneration when intradiscal pressure drops below blood pressure<sup>47</sup>, and the relationship with low back pain is clearest<sup>10</sup>. Astronauts provide an interesting source for investigating discogenic low back pain, they are healthy but immediately upon spaceflight and re-entry experience debilitating low back pain<sup>130,131</sup>. Hypothetically, this could be due to straining of the annulus upon unloading in space, and overloading of the endplates upon re-entry. Their high tendency of developing nucleus protrusion upon re-entry at least indicates a very high intradiscal pressure<sup>131,192</sup>, which has been indicated as a source of discogenic pain<sup>190</sup>. Degeneration of the intervertebral disc could also strain the facet joints due to disc height loss<sup>193</sup>. Similarly, a reduction of intradiscal pressure increases the neutral zone<sup>95,97,98</sup>, which transfers the stabilization of the segment from the disc to adjoining ligaments and muscles<sup>194</sup>. Finally, it is important to consider that the speed of progression through the degenerative circle may depend on the original damage, and pain could be related to the speed of progression<sup>31</sup>.

The use of the degenerative circle as a model for intervertebral disc degeneration has several implications for therapeutic intervention. Currently, there is no cure for intervertebral disc disease; it cannot be reversed, and there is no evidence that it can be slowed down. The degenerative circle provides insight into the disease because it shows that all domains of intervertebral disc degeneration are interdependent (Fig. 2). As such, it suggests that therapies

may be more successful if they affect multiple domains of the degenerative circle in order to slow down or reverse the progressive structural failure (a reversal of the arrows in Fig. 2). An example of such a multi-disciplinary intervention would be a cell-loaded, osmotically active nucleus replacement accompanied by patient-specific physiotherapy. That there is intrinsic healing potential becomes more and more clear, as progenitor cell activity remains present in the human nucleus pulposus<sup>74,195,196</sup>. Furthermore, healing potential in the bovine caudal disc has been shown by the application of physiological loads after chemonucleolysis *ex vivo*<sup>24</sup>: after 14 days, proteoglycan content was restored to pre-intervention levels. However, as this was a model of early disc degeneration, the question arises: is there a point of no return? Additionally, as intervertebral disc degeneration usually develops over years, the duration of such therapies should be considered. Extensive coverage of implications for therapy falls outside the scope of this paper, but this example shows the multi-disciplinary challenge that researchers face when tackling intervertebral disc degeneration.

Intervertebral disc degeneration and the degeneration of joint cartilage as seen in osteoarthritis show marked similarities, although they are rarely discussed simultaneously. Striking similarities are seen on plain radiographs: the loss of disc height or joint space; the sclerosis of the subchondral bone; and the development of osteophytes. Furthermore, the extracellular matrix is comprised of similar constituents (but in another ratio), and similar matrix-degrading enzymes are present in the process of degeneration. There are also differences such as the type of forces applied to the matrix, and the absence of synovial fluid in intervertebral discs; however, a similar degenerative circle could be a model for osteoarthritis.

## Conclusion

The degenerative circle provides a comprehensive model for a contemporary view on intervertebral disc degeneration. It includes a catabolic cell response, changed extracellular matrix, and altered biomechanics. Rather than just simplifying the disease, it also illustrates the complexity as all factors are interdependent, which is why intervertebral disc degeneration has often been called multifactorial. It solves some of the controversy surrounding biomechanics, wear and tear, and cellular physiology by pointing out their interdependency and that all can initiate the degenerative process. Thereby, the model explains some of the human epidemiology and the efficacy of animal models. Because all factors are interrelated, it illustrates why intervertebral disc degeneration is hard to halt or reverse. Rather than being the definitive model for intervertebral disc degeneration, the degenerative circle can serve as a backbone to improve scientific discussion and speed-up therapeutic advancement.

## Author contributions

PV: Conception and design, Collection and assembly of data, Analysis and interpretation of the data, Drafting of the article, Final approval.

IK: Conception and design, Analysis and interpretation of the data, Drafting of the article, Final approval.

KE: Conception and design, Critical revision of the article for important intellectual content, Final approval.

RH: Conception and design, Critical revision of the article for important intellectual content, Final approval.

TW: Conception and design, Critical revision of the article for important intellectual content, Final approval.



BR: Conception and design, Critical revision of the article for important intellectual content, Obtaining of funding, Final approval.

JD: Conception and design, Analysis and interpretation of the data, Critical revision of the article for important intellectual content, Final approval.

TH: Conception and design, Analysis and interpretation of the data, Critical revision of the article for important intellectual content, Obtaining of funding, Final approval.

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#### Conflict of interest

None of the authors have any conflict of interest to declare.

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#### References

- Murray CJL, Vos T, Lozano R, Naghavi M, Flaxman AD, Michaud C, et al. Disability-adjusted life years (DALYs) for 291 diseases and injuries in 21 regions, 1990–2010: a systematic analysis for the Global Burden of Disease Study 2010. *Lancet* 2012;380(9859):2197–223. [http://dx.doi.org/10.1016/S0140-6736\(12\)61689-4](http://dx.doi.org/10.1016/S0140-6736(12)61689-4).
- Spine: low back and neck pain. In: Jacobs JJ, Andersson GB, Bell J-E, Weinstein SL, Dormans JP, Gnatz SM, et al, Eds. *United State Bone and Joint Initiative: The Burden of Musculoskeletal Disease in the United States*. 2nd edn. Rosemont, IL: American Academy of Orthopaedic Surgeons; 2011:21–56.
- Lambeek LC, van Tulder MW, Swinkels ICS, Koppes LLJ, Anema JR, van Mechelen W. The trend in total cost of back pain in The Netherlands in the period 2002 to 2007. *Spine (Phila Pa 1976)* 2011;36(13):1050–8. <http://dx.doi.org/10.1097/BRS.0b013e3181e70488>.
- Cheung KMC, Karppinen J, Chan D, Ho DWH, Song Y-Q, Sham P, et al. Prevalence and pattern of lumbar magnetic resonance imaging changes in a population study of one thousand forty-three individuals. *Spine (Phila Pa 1976)* 2009;34(9):934–40. <http://dx.doi.org/10.1097/BRS.0b013e3181a01b3f>.
- Wang Y, Videman T, Battie MC. ISSLS prize winner: lumbar vertebral endplate lesions: associations with disc degeneration and back pain history. *Spine (Phila Pa 1976)* 2012;37(17):1490–6. <http://dx.doi.org/10.1097/BRS.0b013e3182608ac4>.
- Teraguchi M, Yoshimura N, Hashizume H, Muraki S, Yamada H, Minamide A, et al. Prevalence and distribution of intervertebral disc degeneration over the entire spine in a population-based cohort: the Wakayama Spine Study. *Osteoarthritis Cartilage* 2014;22(1):104–10. <http://dx.doi.org/10.1016/j.joca.2013.10.019>.
- Luoma K, Riihimäki H, Luukkainen R, Raininko R, Viikari-Juntura E, Lamminen A. Low back pain in relation to lumbar disc degeneration. *Spine (Phila Pa 1976)* 2000;25(4):487–92.
- Scheele J, De Schepper EIT, Van Meurs JBJ, Hofman A, Koes BW, Luijsterburg PAJ, et al. Association between spinal morning stiffness and lumbar disc degeneration: the Rotterdam Study. *Osteoarthritis Cartilage* 2012;20(9):982–7. <http://dx.doi.org/10.1016/j.joca.2012.05.011>.
- de Schepper EIT, Damen J, van Meurs JBJ, Ginai AZ, Popham M, Hofman A, et al. The association between lumbar disc degeneration and low back pain: the influence of age, gender, and individual radiographic features. *Spine (Phila Pa 1976)* 2010;35(5):531–6. <http://dx.doi.org/10.1097/BRS.0b013e3181a5b333>.
- Livshits G, Popham M, Malkin I, Sambrook PN, Macgregor AJ, Spector T, et al. Lumbar disc degeneration and genetic factors are the main risk factors for low back pain in women: the UK Twin Spine Study. *Ann Rheum Dis* 2011;70(10):1740–5. <http://dx.doi.org/10.1136/ard.2010.137836>.
- Adams MA, Dolan P, McNally DS. The internal mechanical functioning of intervertebral discs and articular cartilage, and its relevance to matrix biology. *Matrix Biol* 2009;28(7):384–9. <http://dx.doi.org/10.1016/j.matbio.2009.06.004>.
- Adams MA, Freeman BJ, Morrison HP, Nelson IW, Dolan P. Mechanical initiation of intervertebral disc degeneration. *Spine (Phila Pa 1976)* 2000;25(13):1625–36.
- Adams MA, Lama P, Zehra U, Dolan P. Why do some intervertebral discs degenerate, when others (in the same spine) do not? *Clin Anat* 2015;28(2):195–204. <http://dx.doi.org/10.1002/ca.22404>.
- Boubriak OA, Watson N, Sivan SS, Stubbens N, Urban JPG. Factors regulating viable cell density in the intervertebral disc: blood supply in relation to disc height. *J Anat* 2013;222(3):341–8. <http://dx.doi.org/10.1111/joa.12022>.
- Urban JPG, Roberts S. Degeneration of the intervertebral disc. *Arthritis Res Ther* 2003;5(3):120–30. <http://dx.doi.org/10.1186/ar629>.
- Urban JPG, Smith S, Fairbank JCT. Nutrition of the intervertebral disc. *Spine (Phila Pa 1976)* 2004;29(23):2700–9.
- Urban JPG. The role of the physicochemical environment in determining disc cell behaviour. *Biochem Soc Trans* 2002;30(Pt 6):858–64. <http://dx.doi.org/10.1042/>.
- Alpantaki K, Katonis P, Hadjipavlou AG, Spandidos DA, Sourvinos G. Herpes virus infection can cause intervertebral disc degeneration: a causal relationship? *J Bone Joint Surg Br* 2011;93(9):1253–8. <http://dx.doi.org/10.1302/0301-620X.93B9.27002>.
- Hueter C. Anatomische Studien an den Extremitätengelenken Neugeborener und Erwachsener. *Arch für Pathol Anat und Physiol und für Klin Med* 1862;25(5–6):572–99. <http://dx.doi.org/10.1007/BF01879806>.
- Volkman R. Chirurgische Erfahrungen über Knochenverbiegungen und Knochenwachstum. *Arch für Pathol Anat und Physiol und für Klin Med* 1862;24(5–6):512–40. <http://dx.doi.org/10.1007/BF01879454>.
- van der Meulen MCH, Huiskes R. Why mechanobiology? A survey article. *J Biomech* 2002;35(4):401–14.
- Paul CPL, Zuiderbaan HA, Zandieh Doulabi B, van der Veen AJ, van de Ven PM, Smit TH, et al. Simulated-physiological loading conditions preserve biological and mechanical properties of caprine lumbar intervertebral discs in ex vivo culture. *PLoS One* 2012;7(3):e33147. <http://dx.doi.org/10.1371/journal.pone.0033147>.
- Paul CPL, Schoorl T, Zuiderbaan HA, Zandieh Doulabi B, van der Veen AJ, van de Ven PM, et al. Dynamic and static

- overloading induce early degenerative processes in caprine lumbar intervertebral discs. *PLoS One* 2013;8(4):e62411. <http://dx.doi.org/10.1371/journal.pone.0062411>.
24. Gawri R, Moir J, Ouellet J, Beckman L, Steffen T, Roughley P, et al. Physiological loading can restore the proteoglycan content in a model of early IVD degeneration. *PLoS One* 2014;9(7):e101233. <http://dx.doi.org/10.1371/journal.pone.0101233>.
  25. Hsieh AH, Twomey JD. Cellular mechanobiology of the intervertebral disc: new directions and approaches. *J Biomech* 2010;43(1):137–45. <http://dx.doi.org/10.1016/j.jbiomech.2009.09.019>.
  26. Wuertz K, Godburn K, MacLean JJ, Barbir A, Stinnet Donnelly J, Roughley PJ, et al. In vivo remodeling of intervertebral discs in response to short- and long-term dynamic compression. *J Orthop Res* 2009;27(9):1235–42. <http://dx.doi.org/10.1002/jor.20867>.
  27. Setton LA, Chen J. Cell mechanics and mechanobiology in the intervertebral disc. *Spine (Phila Pa 1976)* 2004;29(23):2710–23.
  28. Chan SCW, Ferguson SJ, Gantenbein-Ritter B. The effects of dynamic loading on the intervertebral disc. *Eur Spine J* 2011;20(11):1796–812. <http://dx.doi.org/10.1007/s00586-011-1827-1>.
  29. Lotz JC, Staples A, Walsh A, Hsieh AH. Mechanobiology in intervertebral disc degeneration and regeneration. *Conf Proc IEEE Eng Med Biol Soc Annu Conf* 2004;7:5459. <http://dx.doi.org/10.1109/IEMBS.2004.1404528>.
  30. Adams MA, Roughley PJ. What is intervertebral disc degeneration, and what causes it? *Spine (Phila Pa 1976)* 2006;31(18):2151–61. <http://dx.doi.org/10.1097/01.brs.0000231761.73859.2c>.
  31. Adams MA, Dolan P. Intervertebral disc degeneration: evidence for two distinct phenotypes. *J Anat* 2012;221(6):497–506. <http://dx.doi.org/10.1111/j.1469-7580.2012.01551.x>.
  32. Battié MC, Videman T, Kaprio J, Gibbons LE, Gill K, Manninen H, et al. The Twin Spine Study: contributions to a changing view of disc degeneration. *Spine J* 2009;9(1):47–59. <http://dx.doi.org/10.1016/j.spinee.2008.11.011>.
  33. Battié MC, Videman T. Lumbar disc degeneration: epidemiology and genetics. *J Bone Joint Surg Am* 2006;88(Suppl 2):3–9. <http://dx.doi.org/10.2106/JBJS.E.01313>.
  34. Rajasekaran S, Kanna RM, Senthil N, Raveendran M, Cheung KMC, Chan D, et al. Phenotype variations affect genetic association studies of degenerative disc disease: conclusions of analysis of genetic association of 58 single nucleotide polymorphisms with highly specific phenotypes for disc degeneration in 332 subjects. *Spine J* 2013;23(10):1309–20. <http://dx.doi.org/10.1016/j.spinee.2013.05.019>.
  35. Kepler CK, Ponnappan RK, Tannoury CA, Risbud MV, Anderson DG. The molecular basis of intervertebral disc degeneration. *Spine J* 2013;23(3):318–30. <http://dx.doi.org/10.1016/j.spinee.2012.12.003>.
  36. Weiler C, Nerlich AG, Zipperer J, Bachmeier BE, Boos N. 2002 SSE Award Competition in Basic Science: expression of major matrix metalloproteinases is associated with intervertebral disc degradation and resorption. *Eur Spine J* 2002;11(4):308–20. <http://dx.doi.org/10.1007/s00586-002-0472-0>.
  37. Boos N, Weissbach S, Rohrbach H, Weiler C, Spratt KF, Nerlich AG. Classification of age-related changes in lumbar intervertebral discs: 2002 Volvo Award in basic science. *Spine (Phila Pa 1976)* 2002;27(23):2631–44. <http://dx.doi.org/10.1097/01.BRS.0000035304.27153.5B>.
  38. Zhao C-Q, Wang L-M, Jiang L-S, Dai L-Y. The cell biology of intervertebral disc aging and degeneration. *Ageing Res Rev* 2007;6(3):247–61. <http://dx.doi.org/10.1016/j.arr.2007.08.001>.
  39. Hadjipavlou AG, Tzermiadianos MN, Bogduk N, Zindrick MR. The pathophysiology of disc degeneration: a critical review. *J Bone Joint Surg Br* 2008;90(10):1261–70. <http://dx.doi.org/10.1302/0301-620X.90B10.20910>.
  40. Hsieh AH, Yoon ST. Update on the pathophysiology of degenerative disc disease and new developments in treatment strategies. *Open access J Sport Med* 2010;1:191–9. <http://dx.doi.org/10.2147/OAJSM.S9057>.
  41. Inoue N, Espinoza Orías AA. Biomechanics of intervertebral disk degeneration. *Orthop Clin North Am* 2011;42(4):487–99. <http://dx.doi.org/10.1016/j.jocl.2011.07.001>.
  42. Antoniou J, Steffen T, Nelson F, Winterbottom N, Hollander AP, Poole RA, et al. The human lumbar intervertebral disc: evidence for changes in the biosynthesis and denaturation of the extracellular matrix with growth, maturation, ageing, and degeneration. *J Clin Invest* 1996;98(4):996–1003. <http://dx.doi.org/10.1172/JCI118884>.
  43. Roughley PJ, Melching LI, Heathfield TF, Pearce RH, Mort JS. The structure and degradation of aggrecan in human intervertebral disc. *Eur Spine J* 2006;15(Suppl 3):S326–32. <http://dx.doi.org/10.1007/s00586-006-0127-7>.
  44. Iatridis JC, Nicoll SB, Michalek AJ, Walter BA, Gupta MS. Role of biomechanics in intervertebral disc degeneration and regenerative therapies: what needs repairing in the disc and what are promising biomaterials for its repair? *Spine J* 2013;23(3):243–62. <http://dx.doi.org/10.1016/j.spinee.2012.12.002>.
  45. Brinckmann P, Grootenboer H. Change of disc height, radial disc bulge, and intradiscal pressure from discectomy. An in vitro investigation on human lumbar discs. *Spine (Phila Pa 1976)* 1991;16(6):641–6.
  46. Vergroesen P-PA, van der Veen AJ, van Royen BJ, Kingma I, Smit TH. Intradiscal pressure depends on recent loading and correlates with disc height and compressive stiffness. *Eur Spine J* 2014;23(11):2359–68. <http://dx.doi.org/10.1007/s00586-014-3450-4>.
  47. Sato K, Kikuchi S, Yonezawa T. In vivo intradiscal pressure measurement in healthy individuals and in patients with ongoing back problems. *Spine (Phila Pa 1976)* 1999;24(23):2468–74.
  48. Weiler C, Schietzsch M, Kirchner T, Nerlich AG, Boos N, Wuertz K. Age-related changes in human cervical, thoracic and lumbar intervertebral disc exhibit a strong intra-individual correlation. *Eur Spine J* 2012;21(Suppl 6):S810–8. <http://dx.doi.org/10.1007/s00586-011-1922-3>.
  49. Marchand F, Ahmed AM. Investigation of the laminate structure of lumbar disc annulus fibrosus. *Spine (Phila Pa 1976)* 1990;15(5):402–10.
  50. Skaggs DL, Weidenbaum M, Iatridis JC, Ratcliffe A, Mow VC. Regional variation in tensile properties and biochemical composition of the human lumbar annulus fibrosus. *Spine (Phila Pa 1976)* 1994;19(12):1310–9.
  51. Thompson JP, Pearce RH, Schechter MT, Adams ME, Tsang IK, Bishop PB. Preliminary evaluation of a scheme for grading the gross morphology of the human intervertebral disc. *Spine (Phila Pa 1976)* 1990;15(5):411–5.
  52. Stefanakis M, Luo J, Pollintine P, Dolan P, Adams MA. ISSLS prize winner: mechanical influences in progressive intervertebral disc degeneration. *Spine (Phila Pa 1976)* 2014;39(17):1365–72. <http://dx.doi.org/10.1097/BRS.0000000000000389>.
  53. Holzapfel GA, Schulze-Bauer CAJ, Feigl G, Regitnig P. Single lamellar mechanics of the human lumbar annulus fibrosus. *Biomech Model Mechanobiol* 2005;3(3):125–40. <http://dx.doi.org/10.1007/s10237-004-0053-8>.

54. Ebara S, Iatridis JC, Setton LA, Foster RJ, Mow VC, Weidenbaum M. Tensile properties of nondegenerate human lumbar annulus fibrosus. *Spine (Phila Pa 1976)* 1996;21(4):452–61.
55. Gu WY, Mao XG, Foster RJ, Weidenbaum M, Mow VC, Rawlins BA. The anisotropic hydraulic permeability of human lumbar annulus fibrosus. Influence of age, degeneration, direction, and water content. *Spine (Phila Pa 1976)* 1999;24(23):2449–55.
56. Iatridis JC, Setton LA, Foster RJ, Rawlins BA, Weidenbaum M, Mow VC. Degeneration affects the anisotropic and nonlinear behaviors of human annulus fibrosus in compression. *J Biomech* 1998;31(6):535–44.
57. Rutges JPHJ, Duit RA, Kummer JA, Bekkers JEJ, Oner FC, Castelein RM, et al. A validated new histological classification for intervertebral disc degeneration. *Osteoarthritis Cartilage* 2013;21(12):2039–47. <http://dx.doi.org/10.1016/j.joca.2013.10.001>
58. Wang Y, Videman T, Battié MC. Lumbar vertebral endplate lesions: prevalence, classification, and association with age. *Spine (Phila Pa 1976)* 2012;37(17):1432–9. <http://dx.doi.org/10.1097/BRS.0b013e31824dd20a>.
59. Katz ME, Teitelbaum SL, Gilula LA, Resnick D, Katz SJ. Radiologic and pathologic patterns of end-plate-based vertebral sclerosis. *Invest Radiol* 1988;23(6):447–54.
60. Benneker LM, Heini PF, Alini M, Anderson SE, Ito K. 2004 Young Investigator Award Winner: vertebral endplate marrow contact channel occlusions and intervertebral disc degeneration. *Spine (Phila Pa 1976)* 2005;30(2):167–73.
61. Rutges JPHJ, Jagt van der OP, Oner FC, Verbout AJ, Castelein RJM, Kummer JA, et al. Micro-CT quantification of subchondral endplate changes in intervertebral disc degeneration. *Osteoarthritis Cartilage* 2011;19(1):89–95. <http://dx.doi.org/10.1016/j.joca.2010.09.010>.
62. Cox LGE, van Donkelaar CC, van Rietbergen B, Emans PJ, Ito K. Decreased bone tissue mineralization can partly explain subchondral sclerosis observed in osteoarthritis. *Bone* 2012;50(5):1152–61. <http://dx.doi.org/10.1016/j.bone.2012.01.024>.
63. van Dieën JH, Weinans H, Toussaint HM. Fractures of the lumbar vertebral endplate in the etiology of low back pain: a hypothesis on the causative role of spinal compression in a specific low back pain. *Med Hypotheses* 1999;53(3):246–52. <http://dx.doi.org/10.1054/mehy.1998.0754>.
64. Przybyla A, Pollintine P, Bedzinski R, Adams MA. Outer annulus tears have less effect than endplate fracture on stress distributions inside intervertebral discs: relevance to disc degeneration. *Clin Biomech (Bristol, Avon)* 2006;21(10):1013–9. <http://dx.doi.org/10.1016/j.clinbiomech.2006.07.003>.
65. Roberts S, Urban JP, Evans H, Eisenstein SM. Transport properties of the human cartilage endplate in relation to its composition and calcification. *Spine (Phila Pa 1976)* 1996;21(4):415–20.
66. Mok FPS, Samartzis D, Karppinen J, Luk KDK, Fong DYT, Cheung KMC. ISSLS prize winner: prevalence, determinants, and association of Schmorl nodes of the lumbar spine with disc degeneration: a population-based study of 2449 individuals. *Spine (Phila Pa 1976)* 2010;35(21):1944–52. <http://dx.doi.org/10.1097/BRS.0b013e3181d534f3>.
67. Lotz JC, Fields AJ, Liebenberg EC. The role of the vertebral end plate in low back pain. *Glob Spine J* 2013;3(3):153–64. <http://dx.doi.org/10.1055/s-0033-1347298>.
68. Pfirrmann CW, Metzger A, Zanetti M, Hodler J, Boos N. Magnetic resonance classification of lumbar intervertebral disc degeneration. *Spine (Phila Pa 1976)* 2001;26(17):1873–8.
69. Hunter CJ, Matyas JR, Duncan NA. The notochordal cell in the nucleus pulposus: a review in the context of tissue engineering. *Tissue Eng* 2003;9(4):667–77. <http://dx.doi.org/10.1089/107632703768247368>.
70. Hayes AJ, Benjamin M, Ralphs JR. Role of actin stress fibres in the development of the intervertebral disc: cytoskeletal control of extracellular matrix assembly. *Dev Dyn* 1999;215(3):179–89. 3<179::AID-AJA1>3.0.CO;2-Q. [http://dx.doi.org/10.1002/\(SICI\)1097-0177\(199907\)215](http://dx.doi.org/10.1002/(SICI)1097-0177(199907)215).
71. McCann MR, Tamplin OJ, Rossant J, Seguin CA. Tracing notochord-derived cells using a Noto-cre mouse: implications for intervertebral disc development. *Dis Model Mech* 2012;5:73–82. <http://dx.doi.org/10.1242/dmm.008128>.
72. Choi KS, Cohn MJ, Harfe BD. Identification of nucleus pulposus precursor cells and notochordal remnants in the mouse: Implications for disk degeneration and chordoma formation. *Dev Dyn* 2008;237 (November):3953–8. <http://dx.doi.org/10.1002/dvdy.21805>.
73. Urban JPG. The nucleus of the intervertebral disc from development to degeneration. *Integr Comp Biol* 2000;40(1):53–61. <http://dx.doi.org/10.1093/icb/40.1.53>.
74. Sakai D, Nakamura Y, Nakai T, Mishima T, Kato S, Grad S, et al. Exhaustion of nucleus pulposus progenitor cells with ageing and degeneration of the intervertebral disc. *Nat Commun* 2012;3 (May):1264. <http://dx.doi.org/10.1038/ncomms2226>.
75. Richardson SM, Doyle P, Minogue BM, Gnanalingham K, Hoyland JA. Increased expression of matrix metalloproteinase-10, nerve growth factor and substance P in the painful degenerate intervertebral disc. *Arthritis Res Ther* 2009;11(4):R126. <http://dx.doi.org/10.1186/ar2793>.
76. Purmessur D, Walter BA, Roughley PJ, Laudier DM, Hecht AC, Iatridis J. A role for TNF $\alpha$  in intervertebral disc degeneration: a non-recoverable catabolic shift. *Biochem Biophys Res Commun* 2013;433(1):151–6. <http://dx.doi.org/10.1016/j.bbrc.2013.02.034>.
77. Le Maitre CL, Freemont AJ, Hoyland JA. The role of interleukin-1 in the pathogenesis of human intervertebral disc degeneration. *Arthritis Res Ther* 2005;7(4):R732–45. <http://dx.doi.org/10.1186/ar1732>.
78. Binch A, Cole AA, Breakwell LM, Michael A, Chiverton N, Cross AK, et al. Expression and regulation of neurotrophic and angiogenic factors during human intervertebral disc degeneration. *Arthritis Res Ther* 2014;16(5):416. <http://dx.doi.org/10.1186/s13075-014-0416-1>.
79. V Vo N, Hartman RA, Yurube T, Jacobs LJ, Sowa GA, Kang JD. Expression and regulation of metalloproteinases and their inhibitors in intervertebral disc aging and degeneration. *Spine J* 2013;13(3):331–41. <http://dx.doi.org/10.1016/j.spinee.2012.02.027>.
80. Roughley PJ, Geng Y, Mort JS. The non-aggregated aggrecan in the human intervertebral disc can arise by a non-proteolytic mechanism. *Eur Cell Mater* 2014;28:129–36. discussion 136.
81. Sztrolovics R, Alini M, Roughley PJ, Mort JS. Aggrecan degradation in human intervertebral disc and articular cartilage. *Biochem J* 1997;326(pt 1):235–41.
82. Carter DR, Wong M. Modelling cartilage mechanobiology. *Philos Trans R Soc Lond B Biol Sci* 2003;358(1437):1461–71. <http://dx.doi.org/10.1098/rstb.2003.1346>.
83. Ishihara H, Warensjo K, Roberts S, Urban JP. Proteoglycan synthesis in the intervertebral disk nucleus: the role of extracellular osmolality. *Am J Physiol* 1997;272(5 Pt 1):C1499–506.
84. Iatridis JC, Godburn K, Wuertz K, Alini M, Roughley PJ. Region-dependent aggrecan degradation patterns in the rat



- intervertebral disc are affected by mechanical loading in vivo. *Spine (Phila Pa 1976)* 2011;36(3):203–9. <http://dx.doi.org/10.1097/BRS.0b013e3181cec247>.
85. Jim B, Steffen T, Moir J, Roughley P, Haglund L. Development of an intact intervertebral disc organ culture system in which degeneration can be induced as a prelude to studying repair potential. *Eur Spine J* 2011;20(8):1244–54. <http://dx.doi.org/10.1007/s00586-011-1721-x>.
  86. Wilke HJ, Neef P, Caimi M, Hoogland T, Claes LE. New in vivo measurements of pressures in the intervertebral disc in daily life. *Spine (Phila Pa 1976)* 1999;24(8):755–62.
  87. Nachemson A. The load on lumbar disks in different positions of the body. *Clin Orthop Relat Res* 1966;45:107–22.
  88. Chan WCW, Sze KL, Samartzis D, Leung VYL, Chan D. Structure and biology of the intervertebral disc in health and disease. *Orthop Clin North Am* 2011;42(4):447–64. <http://dx.doi.org/10.1016/j.joc.2011.07.012>.
  89. Urban JP, McMullin JF. Swelling pressure of the lumbar intervertebral discs: influence of age, spinal level, composition, and degeneration. *Spine (Phila Pa 1976)* 1988;13(2):179–87.
  90. Hwang D, Gabai AS, Yu M, Yew AG, Hsieh AH. Role of load history in intervertebral disc mechanics and intradiscal pressure generation. *Biomech Model Mechanobiol* 2012;11(1–2):95–106. <http://dx.doi.org/10.1007/s10237-011-0295-1>.
  91. Lee H-Y, Han L, Roughley PJ, Grodzinsky AJ, Ortiz C. Age-related nanostructural and nanomechanical changes of individual human cartilage aggrecan monomers and their glycosaminoglycan side chains. *J Struct Biol* 2013;181(3):264–73. <http://dx.doi.org/10.1016/j.jsb.2012.12.008>.
  92. Masuoka K, Michalek AJ, MacLean JJ, Stokes IAF, Iatridis JC. Different effects of static versus cyclic compressive loading on rat intervertebral disc height and water loss in vitro. *Spine (Phila Pa 1976)* 2007;32(18):1974–9. <http://dx.doi.org/10.1097/BRS.0b013e3181333d591>.
  93. Adams MA, McMillan DW, Green TP, Dolan P. Sustained loading generates stress concentrations in lumbar intervertebral discs. *Spine (Phila Pa 1976)* 1996;21(4):434–8.
  94. Panjabi MM. Clinical spinal instability and low back pain. *J Electromyogr Kinesiol* 2003;13(4):371–9. [http://dx.doi.org/10.1016/S1050-6411\(03\)00044-0](http://dx.doi.org/10.1016/S1050-6411(03)00044-0).
  95. Zirbel SA, Stolworthy DK, Howell LL, Bowden AE. Intervertebral disc degeneration alters lumbar spine segmental stiffness in all modes of loading under a compressive follower load. *Spine J* 2013;13(9):1134–47. <http://dx.doi.org/10.1016/j.spinee.2013.02.010>.
  96. Ellingson AM, Mehta H, Polly DW, Ellermann J, Nuckley DJ. Disc degeneration assessed by quantitative T2\* (T2 star) correlated with functional lumbar mechanics. *Spine (Phila Pa 1976)* 2013;38(24):E1533–40. <http://dx.doi.org/10.1097/BRS.0b013e3182a59453>.
  97. Quint U, Wilke H-J. Grading of degenerative disk disease and functional impairment: imaging versus patho-anatomical findings. *Eur Spine J* 2008;17(12):1705–13. <http://dx.doi.org/10.1007/s00586-008-0787-6>.
  98. Galbusera F, van Rijsbergen M, Ito K, Huyghe JM, Brayda-Bruno M, Wilke H-J. Ageing and degenerative changes of the intervertebral disc and their impact on spinal flexibility. *Eur Spine J* 2014;23(Suppl 3):S324–32. <http://dx.doi.org/10.1007/s00586-014-3203-4>.
  99. Wang D-L, Jiang S-D, Dai L-Y. Biologic response of the intervertebral disc to static and dynamic compression in vitro. *Spine (Phila Pa 1976)* 2007;32(23):2521–8. <http://dx.doi.org/10.1097/BRS.0b013e318158cb61>.
  100. Haglund L, Moir J, Beckman L, Mulligan KR, Jim B, Ouellet JA, et al. Development of a bioreactor for axially loaded intervertebral disc organ culture. *Tissue Eng Part C Methods* 2011;17(10):1011–9. <http://dx.doi.org/10.1089/ten.TEC.2011.0025>.
  101. Korecki CL, MacLean JJ, Iatridis JC. Dynamic compression effects on intervertebral disc mechanics and biology. *Spine (Phila Pa 1976)* 2008;33(13):1403–9. <http://dx.doi.org/10.1097/BRS.0b013e318175cae7>.
  102. Hartman RA, Bell KM, Debski RE, Kang JD, Sowa GA. Novel ex-vivo mechanobiological intervertebral disc culture system. *J Biomech* 2012;45(2):382–5. <http://dx.doi.org/10.1016/j.jbiomech.2011.10.036>.
  103. Chan SCW, Walser J, Käppeli P, Shamsollahi MJ, Ferguson SJ, Gantenbein-Ritter B. Region specific response of intervertebral disc cells to complex dynamic loading: an organ culture study using a dynamic torsion-compression bioreactor. *PLoS One* 2013;8(8):e72489. <http://dx.doi.org/10.1371/journal.pone.0072489>.
  104. Walsh AJL, Lotz JC. Biological response of the intervertebral disc to dynamic loading. *J Biomech* 2004;37(3):329–37. [http://dx.doi.org/10.1016/S0021-9290\(03\)00290-2](http://dx.doi.org/10.1016/S0021-9290(03)00290-2).
  105. Lee CR, Iatridis JC, Poveda L, Alini M. In vitro organ culture of the bovine intervertebral disc: effects of vertebral endplate and potential for mechanobiology studies. *Spine (Phila Pa 1976)* 2006;31(5):515–22. <http://dx.doi.org/10.1097/01.brs.0000201302.59050.72>.
  106. Handa T, Ishihara H, Ohshima H, Osada R, Tsuji H, Obata K. Effects of hydrostatic pressure on matrix synthesis and matrix metalloproteinase production in the human lumbar intervertebral disc. *Spine (Phila Pa 1976)* 1997;22(10):1085–91.
  107. Ishihara H, McNally DS, Urban JP, Hall AC. Effects of hydrostatic pressure on matrix synthesis in different regions of the intervertebral disc. *J Appl Physiol* 1996;80(3):839–46.
  108. Le Maitre CL, Frain J, Fotheringham AP, Freemont AJ, Hoyland JA. Human cells derived from degenerate intervertebral discs respond differently to those derived from non-degenerate intervertebral discs following application of dynamic hydrostatic pressure. *Biorheology* 2008;45(5):563–75. <http://dx.doi.org/10.3233/BIR-2008-0498>.
  109. van Dijk B, Potier E, Ito K. Culturing bovine nucleus pulposus explants by balancing medium osmolarity. *Tissue Eng Part C Methods* 2011;17(11):1089–96. <http://dx.doi.org/10.1089/ten.TEC.2011.0215>.
  110. Wuertz K, Urban JPG, Klasen J, Ignatius A, Wilke H-J, Claes L, et al. Influence of extracellular osmolarity and mechanical stimulation on gene expression of intervertebral disc cells. *J Orthop Res* 2007;25(11):1513–22. <http://dx.doi.org/10.1002/jor.20436>.
  111. Neidlinger-Wilke C, Mietsch A, Rinkler C, Wilke H-J, Ignatius A, Urban J. Interactions of environmental conditions and mechanical loads have influence on matrix turnover by nucleus pulposus cells. *J Orthop Res* 2012;30(1):112–21. <http://dx.doi.org/10.1002/jor.21481>.
  112. Pritchard S, Erickson GR, Guilak F. Hyperosmotically induced volume change and calcium signaling in intervertebral disc cells: the role of the actin cytoskeleton. *Biophys J* 2002;83(5):2502–10. [http://dx.doi.org/10.1016/S0006-3495\(02\)75261-2](http://dx.doi.org/10.1016/S0006-3495(02)75261-2).
  113. Kim J, Yang S-J, Kim H, Kim Y, Park JB, Dubose C, et al. Effect of shear force on intervertebral disc (IVD) degeneration: an in vivo rat study. *Ann Biomed Eng* 2012;40(9):1996–2004. <http://dx.doi.org/10.1007/s10439-012-0570-z>.
  114. Smith RL, Carter DR, Schurman DJ. Pressure and shear differentially alter human articular chondrocyte metabolism:

- a review. *Clin Orthop Relat Res* 2004;427(Suppl 427): S89–95.
115. Lacroix D, Prendergast PJ. A mechano-regulation model for tissue differentiation during fracture healing: analysis of gap size and loading. *J Biomech* 2002;35(9):1163–1171.
  116. Liu GZ, Ishihara H, Osada R, Kimura T, Tsuji H. Nitric oxide mediates the change of proteoglycan synthesis in the human lumbar intervertebral disc in response to hydrostatic pressure. *Spine (Phila Pa 1976)* 2001;26(2):134–41.
  117. Hsieh AH, Walsh AJL, Cheng LY, Lotz JC. Apoptosis corresponds with disc strain environment during dynamic compression. In: *Transactions of the 50th Annual Meeting of the Orthopaedic Research Society* 2004.
  118. Wang P, Yang L, Hsieh AH. Nucleus pulposus cell response to confined and unconfined compression implicates mechanoregulation by fluid shear stress. *Ann Biomed Eng* 2011;39(3): 1101–11. <http://dx.doi.org/10.1007/s10439-010-0221-1>.
  119. Iatridis JC, Weidenbaum M, Setton LA, Mow VC. Is the nucleus pulposus a solid or a fluid? Mechanical behaviors of the nucleus pulposus of the human intervertebral disc. *Spine (Phila Pa 1976)* 1996;21(10):1174–84.
  120. Iatridis JC, Gwynn IAP. Mechanisms for mechanical damage in the intervertebral disc annulus fibrosus. *J Biomech* 2004;37(8):1165–75. <http://dx.doi.org/10.1016/j.jbiomech.2003.12.026>.
  121. Schollum ML, Robertson PA, Broom ND. ISSLS prize winner: microstructure and mechanical disruption of the lumbar disc annulus: part I: a microscopic investigation of the translamellar bridging network. *Spine (Phila Pa 1976)* 2008;33(25):2702–10. <http://dx.doi.org/10.1097/BRS.0b013e31817bb92c>.
  122. Seidler A, Euler U, Bolm-Audorff U, Ellegast R, Grifka J, Haerting J, et al. Physical workload and accelerated occurrence of lumbar spine diseases: risk and rate advancement periods in a German multicenter case-control study. *Scand J Work Environ Health* 2011;37(1):30–6. <http://dx.doi.org/10.5271/sjweh.3121>.
  123. Battié MC, Videman T, Gill K, Moneta GB, Nyman R, Kaprio J, et al. 1991 Volvo Award in clinical sciences. Smoking and lumbar intervertebral disc degeneration: an MRI study of identical twins. *Spine (Phila Pa 1976)* 1991;16(9):1015–21.
  124. Carragee EJ, Don AS, Hurwitz EL, Cuellar JM, Carrino JA, Carrino J, et al. 2009 ISSLS prize winner: does discography cause accelerated progression of degeneration changes in the lumbar disc: a ten-year matched cohort study. *Spine (Phila Pa 1976)* 2009;34(21):2338–45. <http://dx.doi.org/10.1097/BRS.0b013e3181ab5432>.
  125. Bakker EWP, Verhagen AP, van Trijffel E, Lucas C, Koes BW. Spinal mechanical load as a risk factor for low back pain: a systematic review of prospective cohort studies. *Spine (Phila Pa 1976)* 2009;34(8):E281–93. <http://dx.doi.org/10.1097/BRS.0b013e318195b257>.
  126. Roy TC, Lopez HP, Piva SR. Loads worn by soldiers predict episodes of low back pain during deployment to Afghanistan. *Spine (Phila Pa 1976)* 2013;38(15):1310–7. <http://dx.doi.org/10.1097/BRS.0b013e31829265c4>.
  127. Sward L, Hellström M, Jacobsson B, Nyman R, Peterson L. Disc degeneration and associated abnormalities of the spine in elite gymnasts. A magnetic resonance imaging study. *Spine (Phila Pa 1976)* 1991;16(4):437–43.
  128. Taunton JE, Ryan MB, Clement DB, McKenzie DC, Lloyd-Smith DR, Zumbo BD. A retrospective case-control analysis of 2002 running injuries. *Br J Sports Med* 2002;36(2):95–101.
  129. Dubravac-Simunjak S, Pecina M, Kuipers H, Moran J, Haspl M. The incidence of injuries in elite junior figure skaters. *Am J Sports Med* 2003;31(4):511–7.
  130. V Sayson J, Hargens AR. Pathophysiology of low back pain during exposure to microgravity. *Aviat Space Environ Med* 2008;79(4):365–73.
  131. Johnston SL, Campbell MR, Scheuring R, Feiveson AH. Risk of herniated nucleus pulposus among U.S. Astronauts. *Aviat Space Environ Med* 2010;81(6):566–74. <http://dx.doi.org/10.3357/ASEM.2427.2010>.
  132. Seidler A, Bergmann A, Jäger M, Ellegast R, Ditchen D, Elsner G, et al. Cumulative occupational lumbar load and lumbar disc disease—results of a German multi-center case-control study (EPILIFT). *BMC Musculoskelet Disord* 2009;10: 48. <http://dx.doi.org/10.1186/1471-2474-10-48>.
  133. Coenen P, Kingma I, Boot CRL, Bongers PM, van Dieën JH. Cumulative mechanical low-back load at work is a determinant of low-back pain. *Occup Environ Med* 2014;71(5): 332–7. <http://dx.doi.org/10.1136/oemed-2013-101862>.
  134. Coenen P, Gouttebauge V, van der Burght ASAM, van Dieën JH, Frings-Dresen MHW, van der Beek AJ, et al. The effect of lifting during work on low back pain: a health impact assessment based on a meta-analysis. *Occup Environ Med* 2014;71(12):871–7. <http://dx.doi.org/10.1136/oemed-2014-102346>.
  135. Holguin N, Martin JT, Elliott DM, Judex S. Low-intensity vibrations partially maintain intervertebral disc mechanics and spinal muscle area during deconditioning. *Spine J* 2013;13(4): 428–36. <http://dx.doi.org/10.1016/j.spinee.2013.01.046>.
  136. Holguin N, Uzer G, Chiang F-P, Rubin C, Judex S. Brief daily exposure to low-intensity vibration mitigates the degradation of the intervertebral disc in a frequency-specific manner. *J Appl Physiol* 2011;111(6):1846–53. <http://dx.doi.org/10.1152/japplphysiol.00846.2011>.
  137. Yurube T, Takada T, Suzuki T, Kakutani K, Maeno K, Doita M, et al. Rat tail static compression model mimics extracellular matrix metabolic imbalances of matrix metalloproteinases, aggrecanases, and tissue inhibitors of metalloproteinases in intervertebral disc degeneration. *Arthritis Res Ther* 2012;14(2):R51. <http://dx.doi.org/10.1186/ar3764>.
  138. Hirata H, Yurube T, Kakutani K, Maeno K, Takada T, Yamamoto J, et al. A rat tail temporary static compression model reproduces different stages of intervertebral disc degeneration with decreased notochordal cell phenotype. *J Orthop Res* 2014;32(3):455–63. <http://dx.doi.org/10.1002/jor.22533>.
  139. Miyagi M, Ishikawa T, Kamoda H, Suzuki M, Murakami K, Shibayama M, et al. ISSLS prize winner: disc dynamic compression in rats produces long-lasting increases in inflammatory mediators in discs and induces long-lasting nerve injury and regeneration of the afferent fibers innervating discs: a pathomechanism for chronic discogenic low back pain. *Spine (Phila Pa 1976)* 2012;37(21):1810–8. <http://dx.doi.org/10.1097/BRS.0b013e31824ffac6>.
  140. Kroeber MW, Unglaub F, Wang H, Schmid C, Thomsen M, Nerlich A, et al. New in vivo animal model to create intervertebral disc degeneration and to investigate the effects of therapeutic strategies to stimulate disc regeneration. *Spine (Phila Pa 1976)* 2002;27(23):2684–90. <http://dx.doi.org/10.1097/01.BRS.0000035265.07612.54>.
  141. Court C, Colliou OK, Chin JR, Liebenberg E, Bradford DS, Lotz JC. The effect of static in vivo bending on the murine intervertebral disc. *Spine J* 2001;11(4):239–45.
  142. Court C, Chin JR, Liebenberg E, Colliou OK, Lotz JC. Biological and mechanical consequences of transient intervertebral disc bending. *Eur Spine J* 2007;16(11):1899–906. <http://dx.doi.org/10.1007/s00586-007-0476-x>.
  143. Stokes IAF, McBride C, Aronsson DD, Roughley PJ. Metabolic effects of angulation, compression and reduced mobility on



- annulus fibrosis in a model of altered mechanical environment in scoliosis. *Spine Deform* 2013;1(3):161–70. <http://dx.doi.org/10.1016/j.jspd.2013.02.001>.
144. Bailey JF, Hargens AR, Cheng KK, Lotz JC. Effect of microgravity on the biomechanical properties of lumbar and caudal intervertebral discs in mice. *J Biomech* 2014;47(12):2983–8. <http://dx.doi.org/10.1016/j.jbiomech.2014.07.005>.
  145. Pedrini-Mille A, Maynard JA, Durnova GN, Kaplansky AS, Pedrini VA, Chung CB, et al. Effects of microgravity on the composition of the intervertebral disk. *J Appl Physiol* 1992;73(Suppl 2):26S–32S.
  146. Rajasekaran S, Babu JN, Arun R, Armstrong BRW, Shetty AP, Murugan S. ISSLS prize winner: a study of diffusion in human lumbar discs: a serial magnetic resonance imaging study documenting the influence of the endplate on diffusion in normal and degenerate discs. *Spine (Phila Pa 1976)* 2004;29(23):2654–67.
  147. Lotan R, Oron A, Anekstein Y, Shalmon E, Mirovsky Y. Lumbar stenosis and systemic diseases: is there any relevance? *J Spinal Disord Tech* 2008;21(4):247–51. <http://dx.doi.org/10.1097/BSD.0b013e31813707af>.
  148. Devaraj S, Cheung AT, Jialal I, Griffen SC, Nguyen D, Glaser N, et al. Evidence of increased inflammation and microcirculatory abnormalities in patients with type 1 diabetes and their role in microvascular complications. *Diabetes* 2007;56(11):2790–6. <http://dx.doi.org/10.2337/db07-0784>.
  149. Sørensen LT, Jørgensen S, Petersen LJ, Hemmingsen U, Bülow J, Loft S, et al. Acute effects of nicotine and smoking on blood flow, tissue oxygen, and aerobic metabolism of the skin and subcutis. *J Surg Res* 2009;152(2):224–30. <http://dx.doi.org/10.1016/j.jss.2008.02.066>.
  150. Pellaton C, Kubli S, Feihl F, Waeber B. Blunted vasodilatory responses in the cutaneous microcirculation of cigarette smokers. *Am Heart J* 2002;144(2):269–74. <http://dx.doi.org/10.1067/mhj.2002.123842>.
  151. Chen S, Liao M, Li J, Peng H, Xiong M. The correlation between microvessel pathological changes of the endplate and degeneration of the intervertebral disc in diabetic rats. *Exp Ther Med* 2013;5(3):711–7. <http://dx.doi.org/10.3892/etm.2012.868>.
  152. Albert HB, Sorensen JS, Christensen BS, Manniche C. Antibiotic treatment in patients with chronic low back pain and vertebral bone edema (Modic type 1 changes): a double-blind randomized clinical controlled trial of efficacy. *Eur Spine J* 2013;22(4):697–707. <http://dx.doi.org/10.1007/s00586-013-2675-y>.
  153. Carter JD, Gerard HC, Whittum-Hudson JA, Hudson AP. The molecular basis for disease phenotype in chronic Chlamydia-induced arthritis. *Int J Clin Rheumatol* 2012;7(6):627–40. <http://dx.doi.org/10.2217/ijr.12.65>.
  154. Zeidler H, Hudson AP. New insights into Chlamydia and arthritis. Promise of a cure? *Ann Rheum Dis* 2014;73(4):637–44. <http://dx.doi.org/10.1136/annrheumdis-2013-204110>.
  155. Alvarez-Lafuente R, Fernández-Gutiérrez B, de Miguel S, Jover JA, Rollin R, Loza E, et al. Potential relationship between herpes viruses and rheumatoid arthritis: analysis with quantitative real time polymerase chain reaction. *Ann Rheum Dis* 2005;64(9):1357–9. <http://dx.doi.org/10.1136/ard.2004.033514>.
  156. Wang D, Nasto LA, Roughley P, Leme AS, Houghton AM, Usas A, et al. Spine degeneration in a murine model of chronic human tobacco smokers. *Osteoarthritis Cartilage* 2012;20(8):896–905. <http://dx.doi.org/10.1016/j.joca.2012.04.010>.
  157. Nasto LA, Wang D, Robinson AR, Clauson CL, Ngo K, Dong Q, et al. Genotoxic stress accelerates age-associated degenerative changes in intervertebral discs. *Mech Ageing Dev* 2013;134(1–2):35–42. <http://dx.doi.org/10.1016/j.mad.2012.11.002>.
  158. Adler JH, Schoenbaum M, Silberberg R. Early onset of disk degeneration and spondylosis in sand rats (*Psammomys obesus*). *Vet Pathol* 1983;20(1):13–22.
  159. Tsai T-T, Ho NY-J, Lin Y-T, Lai P-L, Fu T-S, Niu C-C, et al. Advanced glycation end products in degenerative nucleus pulposus with diabetes. *J Orthop Res* 2014;32(2):238–44. <http://dx.doi.org/10.1002/jor.22508>.
  160. Nasto LA, Ngo K, Leme AS, Robinson AR, Dong Q, Roughley P, et al. Investigating the role of DNA damage in tobacco smoking-induced spine degeneration. *Spine J* 2014;24(3):416–23. <http://dx.doi.org/10.1016/j.spinee.2013.08.034>.
  161. Won H-Y, Park J-B, Park E-Y, Riew KD. Effect of hyperglycemia on apoptosis of notochordal cells and intervertebral disc degeneration in diabetic rats. *J Neurosurg Spine* 2009;11(6):741–8. <http://dx.doi.org/10.3171/2009.6.SPINE09198>.
  162. Park E-Y, Park J-B. Dose- and time-dependent effect of high glucose concentration on viability of notochordal cells and expression of matrix degrading and fibrotic enzymes. *Int Orthop* 2013;37(6):1179–86. <http://dx.doi.org/10.1007/s00264-013-1836-2>.
  163. Phillips KLE, Jordan-Mahy N, Nicklin MJH, Le Maitre CL. Interleukin-1 receptor antagonist deficient mice provide insights into pathogenesis of human intervertebral disc degeneration. *Ann Rheum Dis* 2013;72(11):1860–7. <http://dx.doi.org/10.1136/annrheumdis-2012-202266>.
  164. Furtwängler T, Chan SCW, Bahrenberg G, Richards PJ, Gantenbein-Ritter B. Assessment of the matrix degenerative effects of MMP-3, ADAMTS-4, and HTRA1, injected into a bovine intervertebral disc organ culture model. *Spine (Phila Pa 1976)* 2013;38(22):E1377–87. <http://dx.doi.org/10.1097/BRS.0b013e31829ffde8>.
  165. Lama P, Le Maitre CL, Dolan P, Tarlton JF, Harding IJ, Adams MA. Do intervertebral discs degenerate before they herniate, or after? *Bone Joint J* 2013;95-B(8):1127–33. <http://dx.doi.org/10.1302/0301-620X.95B8.31660>.
  166. Dudli S, Ferguson SJ, Haschtmann D. Severity and pattern of post-traumatic intervertebral disc degeneration depend on the type of injury. *Spine J* 2014;24(7):1256–64. <http://dx.doi.org/10.1016/j.spinee.2013.07.488>.
  167. Alkhatib B, Rosenzweig DH, Krock E, Roughley PJ, Beckman L, Steffen T, et al. Acute mechanical injury of the human intervertebral disc: link to degeneration and pain. *Eur Cell Mater* 2014;28(514):98–110. discussion 110–1.
  168. Yoganandan N, Larson SJ, Pintar FA, Gallagher M, Reinartz J, Droese K. Intravertebral pressure changes caused by spinal microtrauma. *Neurosurgery* 1994;35(3):415–21. discussion 421.
  169. Holm S, Holm AK, Ekström L, Karladani A, Hansson T. Experimental disc degeneration due to endplate injury. *J Spinal Disord Tech* 2004;17(1):64–71.
  170. Dudli S, Haschtmann D, Ferguson SJ. Fracture of the vertebral endplates, but not equine energetic impact load, promotes disc degeneration in vitro. *J Orthop Res* 2012;30(5):809–16. <http://dx.doi.org/10.1002/jor.21573>.
  171. Quero L, Klawitter M, Schmaus A, Rothley M, Sleeman J, Tiaden AN, et al. Hyaluronic acid fragments enhance the inflammatory and catabolic response in human intervertebral disc cells through modulation of toll-like receptor 2 signaling pathways. *Arthritis Res Ther* 2013;15(4):R94. <http://dx.doi.org/10.1186/ar4274>.
  172. Fry TR, Eurell JC, Johnson AL, Brown MD, Losonsky JM, Schaeffer DJ. Radiographic and histologic effects of

- chondroitinase ABC on normal canine lumbar intervertebral disc. *Spine (Phila Pa 1976)* 1991;16(7):816–9.
173. Norcross JP, Lester GE, Weinhold P, Dahners LE. An in vivo model of degenerative disc disease. *J Orthop Res* 2003;21(1):183–8. [http://dx.doi.org/10.1016/S0736-0266\(02\)00098-0](http://dx.doi.org/10.1016/S0736-0266(02)00098-0).
  174. Hoogendoorn RJ, Wuisman PI, Smit TH, Everts VE, Helder MN. Experimental intervertebral disc degeneration induced by chondroitinase ABC in the goat. *Spine (Phila Pa 1976)* 2007;32(17):1816–25. <http://dx.doi.org/10.1097/BRS.0b013e31811ebac5>.
  175. Sasaki M, Takahashi T, Miyahara K, Hirose aT. Effects of chondroitinase ABC on intradiscal pressure in sheep: an in vivo study. *Spine (Phila Pa 1976)* 2001;26(5):463–8.
  176. Masuda K, Aota Y, Muehleman C, Imai Y, Okuma M, Thonar EJ, et al. A novel rabbit model of mild, reproducible disc degeneration by an anulus needle puncture: correlation between the degree of disc injury and radiological and histological appearances of disc degeneration. *Spine (Phila Pa 1976)* 2005;30(1):5–14.
  177. Zhang H, La Marca F, Hollister SJ, Goldstein SA, Lin C-Y. Developing consistently reproducible intervertebral disc degeneration at rat caudal spine by using needle puncture. *J Neurosurg Spine* 2009;10(6):522–30. <http://dx.doi.org/10.3171/2009.2.SPINE08925>.
  178. Han B, Zhu K, Li F-C, Xiao Y-X, Feng J, Shi Z-L, et al. A simple disc degeneration model induced by percutaneous needle puncture in the rat tail. *Spine (Phila Pa 1976)* 2008;33(18):1925–34. <http://dx.doi.org/10.1097/BRS.0b013e31817c64a9>.
  179. Kong MH, Do DH, Miyazaki M, Wei F, Yoon S, Wang JC. Rabbit model for in vivo study of intervertebral disc degeneration and regeneration. *J Korean Neurosurg Soc* 2008;44(5):327–33. <http://dx.doi.org/10.3340/jkns.2008.44.5.327>.
  180. Vadalà G, De Strobel F, Bernardini M, Denaro L, D'Avella D, Denaro V. The transpedicular approach for the study of intervertebral disc regeneration strategies: in vivo characterization. *Eur Spine J* 2013;22(Suppl 6):S972–8. <http://dx.doi.org/10.1007/s00586-013-3007-y>.
  181. Holm S, Ekström L, Holm AK, Hansson T. Intradiscal pressure in the degenerated porcine intervertebral disc. *Vet Comp Orthop Traumatol* 2007;20(1):29–33.
  182. Haschtmann D, Stoyanov JV, Gédet P, Ferguson SJ. Vertebral endplate trauma induces disc cell apoptosis and promotes organ degeneration in vitro. *Eur Spine J* 2008;17(2):289–99. <http://dx.doi.org/10.1007/s00586-007-0509-5>.
  183. Holm S, Mackiewicz Z, Holm AK, Konttinen YT, Kouri V-P, Indahl A, et al. Pro-inflammatory, pleiotropic, and anti-inflammatory TNF-alpha, IL-6, and IL-10 in experimental porcine intervertebral disk degeneration. *Vet Pathol* 2009;46(6):1292–300. <http://dx.doi.org/10.1354/vp.07-VP-0179-K-FL>.
  184. Colombini A, Lombardi G, Corsi MM, Banfi G. Pathophysiology of the human intervertebral disc. *Int J Biochem Cell Biol* 2008;40(5):837–42. <http://dx.doi.org/10.1016/j.biocel.2007.12.011>.
  185. Arun R, Freeman BJC, Scammell BE, McNally DS, Cox E, Gowland P. 2009 ISSLS Prize Winner: what influence does sustained mechanical load have on diffusion in the human intervertebral disc?: an in vivo study using serial post-contrast magnetic resonance imaging. *Spine (Phila Pa 1976)* 2009;34(21):2324–37. <http://dx.doi.org/10.1097/BRS.0b013e3181b4df92>.
  186. Rajan NE, Bloom O, Maidhof R, Stetson N, Sherry B, Levine M, et al. Toll-Like Receptor 4 (TLR4) expression and stimulation in a model of intervertebral disc inflammation and degeneration. *Spine (Phila Pa 1976)* 2013;38(16):1343–51. <http://dx.doi.org/10.1097/BRS.0b013e31826b71f4>.
  187. Mayer JE, Iatridis JC, Chan D, Qureshi SA, Gottesman O, Hecht AC. Genetic polymorphisms associated with intervertebral disc degeneration. *Spine J* 2013;13(3):299–317. <http://dx.doi.org/10.1016/j.spinee.2013.01.041>.
  188. Locke AE, Kahali B, Berndt SI, Justice AE, Pers TH, Day FR, et al. Genetic studies of body mass index yield new insights for obesity biology. *Nature* 2015;518(7538):197–206. <http://dx.doi.org/10.1038/nature14177>.
  189. Bergknut N, Rutges JPHJ, Kranenburg H-JC, Smolders LA, Hagman R, Smidt H-J, et al. The dog as an animal model for intervertebral disc degeneration? *Spine (Phila Pa 1976)* 2012;37(5):351–8. <http://dx.doi.org/10.1097/BRS.0b013e31821e5665>.
  190. Bogduk N, Aprill C, Derby R. Lumbar discogenic pain: state-of-the-art review. *Pain Med* 2013;14(6):813–36. <http://dx.doi.org/10.1111/pme.12082>.
  191. Freemont AJ, Watkins A, Le Maitre C, Baird P, Jeziorska M, Knight MTN, et al. Nerve growth factor expression and innervation of the painful intervertebral disc. *J Pathol* 2002;197(3):286–92. <http://dx.doi.org/10.1002/path.1108>.
  192. Wilke H-J, Ressel L, Heuer F, Graf N, Rath S. Can prevention of a reherniation be investigated? Establishment of a herniation model and experiments with an anular closure device. *Spine (Phila Pa 1976)* 2013;38(10):E587–93. <http://dx.doi.org/10.1097/BRS.0b013e31828ca4bc>.
  193. Pollintine P, Przybyla AS, Dolan P, Adams MA. Neural arch load-bearing in old and degenerated spines. *J Biomech* 2004;37(2):197–204. [http://dx.doi.org/10.1016/S0021-9290\(03\)00308-7](http://dx.doi.org/10.1016/S0021-9290(03)00308-7).
  194. Panjabi MM. The stabilizing system of the spine. Part II. Neutral zone and instability hypothesis. *J Spinal Disord* 1992;5(4):390–6. discussion 397.
  195. V Risbud M, Guttapalli A, Tsai T-T, Lee JY, Danielson KG, Vaccaro AR, et al. Evidence for skeletal progenitor cells in the degenerate human intervertebral disc. *Spine (Phila Pa 1976)* 2007;32(23):2537–44. <http://dx.doi.org/10.1097/BRS.0b013e318158dea6>.
  196. van den Akker GG, Surtel DA, Cremers A, Rodrigues-Pinto R, Richardson SM, Hoyland JA, et al. Novel immortal human cell lines reveal subpopulations in the nucleus pulposus. *Arthritis Res Ther* 2014;16(3):R135. <http://dx.doi.org/10.1186/ar4597>.